#### ARTHROPODS AND MEDICAL ENTOMOLOGY - SHORT COMMUNICATION



# Ticks and tick-borne pathogens infecting livestock and dogs in Tchicala-Tcholoanga, Huambo Province, Angola

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#### Abstract

The diversity of ticks and tick-borne pathogens (TBPs) infesting domestic animals in Tchicala-Tcholoanga, Angola, in 2016 was investigated. Seventeen tick species were recorded, *Amblyomma pomposum* being the most abundant on cattle (40%), goats (38%) and sheep (35%); *Rhipicephalus turanicus* was the most abundant on dogs (46%). This study presents new records of *Haemaphysalis paraleachi*, *R. compositus*, *R. kochi* and *R. sulcatus* in Angola, the first georeferenced population of *Ha. leachi* in southern Africa and the second record of *R. microplus* in Angola. Using the reverse line blot (RLB) hybridisation assay, fifteen TBP species were detected in blood samples from cattle (n = 88), goats (n = 82), sheep (n = 85) and dogs (n = 85). F The most frequently detected species were *Theileria velifera* in cattle (78%), *Theileria ovis* in sheep (80%) and *Babesia vogeli* in dogs (35%). Species-specific quantitative PCR assays detected *Babesia bigemina* in 43% (35/80) of blood samples of cattle, while *E. ruminantium* was detected in 4% (3/70) of blood samples and in 7% of *A. pomposum* ticks. *Anaplasma platys* was detected from cattle (18%) and sheep (6%) during RLB analysis. These findings constitute pioneering research in Angola.

Keywords Ticks · Reverse line blot · Anaplasma · Babesia · Theileria · Angola

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# Introduction

Ticks are important pathogen vectors that negatively impact the health of animal and human populations across the world and are considered a major constraint to livestock production (Jongejan and Uilenberg 2004). In Africa, 40 tick species are recognised as a threat to domestic animal health (Walker et al. 2003). Species of major veterinary and economic importance belong to the genera *Amblyomma*, *Hyalomma*, and *Rhipicephalus*, and are commonly associated with the transmission of parasites of the genera *Anaplasma*, *Ehrlichia*, *Babesia* and *Theileria* (Walker et al. 2003). Domestic ruminants and dogs are the most affected by these haemoparasites (Jongejan and Uilenberg 2004).

To understand the current distribution of ticks in Angola, it is important to consider the impact of major historical events, especially the armed conflict that affected the country for over three decades. It is believed that the war led to uncontrolled animal dispersion as well as a drastic reduction in livestock during the 1990s. Consequently, animals were imported from neighbouring countries, namely, Botswana (Gomes 1993), Namibia and Zambia (A. Gomes, personal communication, 2015) and from overseas, particularly Brazil (A. José, personal communication, 2015). The livestock population in Angola comprises mainly cattle and goats, although sheep and pigs can be found in both the traditional and commercial sectors. Dogs are frequently present in grazing areas, where they are used to herd livestock and to hunt. The first studies on ticks in Angola were conducted by Dias (1948) in various regions of the country. Subsequent studies were conducted by Serrano (1963), Dias (1983), and Gomes et al. (1994).

Despite the wide range of molecular tools currently available for the diagnosis of tick-borne diseases (TBDs), classical diagnostic assays, such as blood smears and serological tests, are practically the only methods previously used to detect haemoparasites in Angola. The aim of this study was to update information on the diversity of ticks infesting livestock and dogs as well as tick-borne pathogens detected using DNAbased methods in Angola.

### Material and methods

The fieldwork for this cross-sectional study was conducted in four communes: Mbave (13 sampling sites), Sambo (13 sites), Samboto (11 sites) and Sede (12 sites) in Tchicala-Tcholoanga Municipality, Huambo Province, Angola (Fig. 1). The annual rainfall is 1200–1600 mm during the wet season (October to April), and there is no rainfall during the dry season (May to September) (http://www.angola.climatemps. com/). The annual average temperature is 20 °C. The main economic activity in the municipality is agriculture, and livestock production is based on indigenous breeds reared on small traditional farms.

In January 2016, ticks and blood samples were collected from cattle, goats, sheep, and dogs of various ages and both

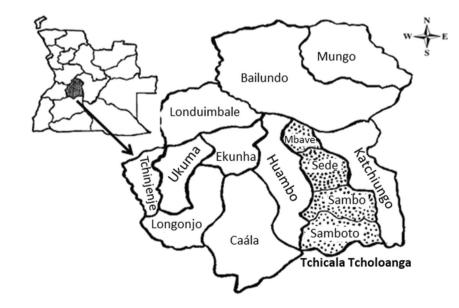
Fig. 1 Map of Huambo Province in Angola showing the 11 municipalities, including Tchicala-Tcholoanga Municipality (shaded with dots), where a study on ticks and tickborne pathogens was conducted. All four communes (Mbave, Sede, Sambo and Samboto) in Tchicala-Tcholoanga Municipality were sampled. Inset is a map of Angola showing the location of Huambo Province sexes in the four communes. The month of January corresponds to the mid-rainy season, which is the peak of adult tick activity for the species occurring in this region. Adult ticks were targeted because they are easier to identify to species level than their immature stages. Given our aim of determining the presence or absence of tick-borne pathogens (rather than prevalence), the required sample size was calculated using the formula below (Thrusfield 2007):

$$n = \left[1 - (1 - P1)^{\frac{1}{d}}\right] \left[N - \frac{d}{2}\right] + 1$$

where *n* = required sample size; P1 = probability of finding at least one case in the sample (95% probability = 0.95); *N* = population size or number of animals for each host species (considered as an infinite population,  $\infty$ ); *d* = minimum number of animals affected in the population. The expected occurrence rate of tick-borne pathogens was assumed to be 15%, derived from experience in our laboratory, in which 15% is one of the lowest percentages of parasite occurrence in endemic areas. Therefore,  $\geq$  19 individuals of each host species were estimated to be sufficient (Cannon and Roe, 1982, cited in Thrusfield 2007) to detect tick-borne pathogens in the study area.

All visible ticks were manually collected from half of the body of five of the most infested animals of each host species in a commune. The total number of ticks collected from an animal represented a single tick collection. The ticks were identified to species level by stereomicroscopic examination using the keys published by Walker et al. (2003).

A total of 340 animals, 88 cattle, 82 goats, 85 sheep and 85 dogs, were sampled and 4 ml of blood were collected from each animal. DNA was extracted from each blood sample



using the QIAamp® DNA Mini Kit (QIAGEN, Hilden, Germany) and then stored at -20 °C. The 340 blood samples were tested using RLB assay as previously described (Nijhof et al. 2005). The positive controls were DNA extracted from *Anaplasma centrale* and *Babesia bovis* live blood vaccines (Onderstepoort Biological Products [OBP], Pretoria, South Africa), for the 16S rRNA and 18S rRNA PCRs, respectively, while nuclease-free water was used as negative control. *Anaplasma, Ehrlichia, Babesia* and *Theileria* genus-specific probes and 36 species-specific probes, as outlined in Byaruhanga et al. (2016), and a probe specific for *A. platys* (Sirigireddy and Ganta 2005) were included on the RLB membrane.

Two quantitative real-time PCR (qPCR) assays were used for detection of B. bigemina and B. bovis DNA from 88 blood samples from cattle, as described previously (Kim et al. 2007), except that the annealing temperatures were adjusted to 57 °C for B. bigemina and 54 °C for B. bovis. These assays used the same negative and positive controls as those described for RLB. A TaqMan qPCR assay was conducted to amplify a 226 bp fragment of the conserved pCS20 region of E. ruminantium (Steyn et al. 2008) from 70 blood samples (cattle: 20; goats: 20; sheep: 20; dogs: 10). One hundred randomly selected Amblyomma pomposum ticks (male and female) collected from apparently healthy ruminants were also analysed. DNA extraction from individual dissected ticks was performed using the DNeasy® Blood & Tissue Kit (QIAGEN, Hilden, Germany), following the manufacturer's protocol. The positive control was E. ruminantium Welgevonden genomic DNA, while nuclease-free water was used as the negative control. The near full-length parasite 16S rRNA gene for Anaplasma/Ehrlichia spp. was amplified, cloned and sequenced from 11 DNA samples (cattle: 6; sheep: 5) that showed A. platys-specific signals during RLB analysis. The amplification and sequence analyses were performed as described previously (Byaruhanga et al. 2018).

## **Results and discussion**

A total of 2963 ticks belonging to five genera and 17 species were collected from domestic animals in Tchicala-Tcholoanga Municipality in Angola (Table 1). In previous surveys in Angola, Serrano (1963) and Dias (1983) recorded 34 and 19 tick species, respectively. Subsequently, Gomes et al. (1994) recovered 13 species in a survey conducted in Huíla Province. *Haemaphysalis paraleachi, R. compositus, R. kochi* and *R. sulcatus* are new records for Angola, and this is the second record for *R. microplus*.

Nine tick species; *A. pomposum, Haemaphysalis leachi, R. decoloratus, R. evertsi mimeticus, R. lunulatus, R. punctatus, R. sulcatus, R. turanicus* and *R. tricuspis* were present in all four communes. Twelve species were recovered in Sambo, 13

in Mbave and 15 species each in Samboto and Sede. There was a generally similar pattern of relative abundance of each tick species in each commune, for each host species. *R. microplus* (10 specimens) was only found in Sede in one sampled bovine.

. Amblyomma pomposum was the most abundant species amongst cattle, sheep and goats (Table 1). Of the 15 species of Amblyomma that have been reported in Angola, A. pomposum is the most common and is regarded as an important vector of E. ruminantium, the cause of heartwater in ruminants (Dias 1948; Serrano 1963; Gomes et al. 1994). Its distribution is restricted to Angola, western Zambia and the southern region of the Democratic Republic of Congo (Walker et al. 2003). Gomes and Neves (2018) found R. microplus in two provinces of Angola, and the present study records its presence on a bovine in a third locality. Additionally, this study reports the first georeferenced population of Ha. leachi in southern Africa since the reinstatement of Ha. elliptica as a valid species . Rhipicephalus turanicus was the most abundant tick species on dogs, and its possible role as a vector of B. vogeli and E. canis should be evaluated.

Using RLB, 15 haemoparasite species were detected from the 340 DNA samples from cattle, goats, sheep and dogs (Table 2). However, genus-specific signals without speciesspecific signals were also observed. Theileria mutans, T. velifera and Theileria sp. (sable) were the most commonly detected species in cattle (> 70% each). Theileria mutans and T. velifera are transmitted by A. hebraeum, A. lepidum and A. variegatum (Walker et al. 2003). Considering that these ticks are absent in the study region, the current findings suggest that A. pomposum is also a vector. Theileria velifera has previously been recorded in Angola (Kubelová et al. 2012). Theileria sp. (sable) is a parasite of sable antelope (Stoltsz and Dunsterville 1992, cited by Nijhof et al. 2005), and reports of widespread occurrence of the parasite in cattle require DNA sequencing studies, to confirm circulation of the pathogen in cattle populations, and to rule out cross-reactions between Theileria sp. (sable) and T. velifera RLB probes. The infection rate of A. marginale was 28% (Table 2), which is lower than that found in Bié Province in Angola (38%) (Kubelová et al. 2012), using conventional PCR. Anaplasma platys which is not a known pathogen of cattle was also detected. Nine haemoparasites were detected in sheep and T. ovis was the most abundant species (80%) (Table 2). Although T. ovis is considered a haemoparasite of negligible pathogenic importance, it was implicated in massive sheep losses in Pakistan (Durrani et al. 2011). It is transmitted by several tick species including Amblyomma spp., Hyalomma spp. and Rhipicephalus spp., all of which were found on sheep in this study. The detection of T. ovis in dogs in the present study was unexpected and, to our knowledge, has not been previously reported. Thus, further investigation is needed. The detection of A. platys, T. bicornis, B. bovis and B. rossi in sheep

 Table 1
 Ticks collected from domestic ruminants and dogs in a cross-sectional study in Tchicala-Tcholoanga, Huambo Province, Angola

Tick species	Cattle $(n = 20)$		Goats $(n = 20)$		Sheep $(n = 20)$		Dogs $(n = 20)$	
	No. of ticks (%)	No. of animals infested (%)	No. of ticks (%)	No. of animals infested (%)	No. of ticks (%)	No. of animals infested (%)	No. of ticks (%)	No. of animals infested (%)
A. pomposum*	617 (40.2)	20 (100)	52 (38.2)	14 (70)	127 (35.4)	16 (80)	8 (0.9)	6 (30)
Ha. Leachi <sup>#</sup>	0 (0.0)	-	0 (0.0)	-	0 (0.0)	-	10 (1.1)	3 (15)
Ha. paraleachi <sup>#</sup>	0 (0.0)	-	0 (0.0)	-	0 (0.0)	-	57 (6.1)	12 (60)
Hy. Truncatum	270 (17.6)	14 (70)	2 (1.5)	1 (5)	27 (7.5)	6 (30)	0 (0.0)	-
I. cavipalpus	7 (0.5)	3 (15)	1 (0.7)	1 (5)	0 (0.0)	-	13 (1.4)	4 (20)
R. compositus	12 (0.8)	5 (25)	0 (0.0)	-	1 (0.3)	1 (5)	0 (0.0)	-
R. decoloratus*	466 (30.4)	18 (90)	3 (2.2)	3 (15)	34 (9.5)	10 (50)	4 (0.4)	2 (10)
R. evertsi evertsi	2 (0.1)	2 (10)	3 (2.2)	2 (10)	8 (2.2)	5 (25)	0 (0.0)	-
R. e. mimeticus	61 (3.9)	10 (50)	16 (11.8)	9 (45)	50 (13.9)	5 (25)	0 (0.0)	-
R. kochi	22 (1.4)	7 (35)	0 (0.0)	-	0 (0.0)	-	0 (0.0)	-
R. lunulatus*	11 (0.7)	4 (20)	4 (2.9)	3 (15)	4 (1.1)	3 (15)	52 (5.6)	16 (80)
R. microplus	10 (0.7)	1 (5)	0 (0.0)	-	0 (0.0)	-	0 (0.0)	-
R. punctatus*	22 (1.4)	7 (35)	41 (30.2)	8 (40)	91 (25.4)	9 (45)	85 (9.1)	9 (45)
R. simus <sup>#</sup>	0 (0.0)	-	0 (0.0)	-	0 (0.0)	-	6 (0.6)	4 (20)
R. sulcatus <sup>#</sup>	0 (0.0)	-	0 (0.0)	-	0 (0.0)	-	84 (9.0)	14 (70)
R. tricuspis*	35 (2.3)	11 (55)	14 (10.3)	8 (40)	17 (4.7)	7 (35)	187 (20.0)	18 (90)
R. turanicus <sup>#</sup>	0 (0.0)	-	0 (0.0)		0 (0.0)	-	427 (45.8)	18 (90)
Total	1535 (100)		136 (100)		359 (100)		933 (100)	

\*Tick species infesting all four animal species; #Tick species that were only collected from dogs

A. = Amblyomma; Ha. = Haemaphysalis; Hy. = Hyalomma; R. = Rhipicephalus; I. = Ixodes

(no.) = number of animals infested or number of ticks collected; Percentage (%) of the total in parenthesis

(Table 2) was unforeseen. Of the 82 blood samples from goats, only two were positive for tick-borne parasites (Table 2). In contrast, 100% of goats examined in the neighbouring Bié province and were infected with *A. ovis* using PCR (Kubelová et al. 2012). Seventy-five (88%) blood

samples from dogs were positive for one or more pathogens. Four haemoparasite species were identified and the most frequently occurring was *B. vogeli* (35%) followed by *B. rossi* (24%). In contrast, Cardoso et al. (2016) reported *A. platys* and *Hepatozoon canis* as the most frequent parasite species in

Table 2Proportions of bloodsamples from domestic ruminantsand dogs from TchicalaTcholohanga municipality,province of Huambo, Angola,found positive for tick-bornepathogens, using the reverse lineblot hybridisation assay

Tick-borne pathogen	Number and percentage of positive samples						
	Cattle $(n = 88)$	Sheep $(n = 85)$	Goats ( <i>n</i> = 82)	Dogs ( <i>n</i> = 85)			
Anaplasma centrale	11 (12.5)	2 (2.4)	2 (2.4)	_			
Anaplasma marginale	25 (28.4)	1 (1.2)	-	-			
Anaplasma bovis	1 (1.1)	-	-	-			
Anaplasma platys	16 (18.2)	5 (5.9)	-	-			
Anaplasma sp. Omatjenne	22 (25)	4 (4.7)	-	-			
Ehrlichia canis	-	-	-	1 (1.2)			
Babesia bigemina	3 (3.4)	-	-	-			
Babesia bovis	-	1 (1.2)	-	-			
Babesia rossi	-	1 (1.2)	-	20 (23.5)			
Babesia vogeli	-	-	-	30 (35.3)			
Theileria sp. (sable)	63 (71.6)	46 (54.1)	-	-			
Theileria mutans	65 (73.9)	-	-	-			
Theileria velifera	69 (78.4)	-	-	-			
Theileria bicornis	-	4 (4.7)	-	-			
Theileria ovis	-	68 (80)	-	7 (8.2)			

dogs in Luanda. Furthermore, infection rates with *B. vogeli* (35%) in our study were considerably higher than those recorded by Cardoso et al. (2016) in Luanda (5.8%). The detection of both *E. canis* and *B. vogeli* in dogs, in the absence of *R. sanguineus*, indicates a potential vector role for *R. turanicus*. However, our findings do not exclude the possibility of the presence of *R. sanguineus*. *Babesia rossi* is known to be transmitted by *Ha. elliptica*. In this study, the occurrence of *B. rossi* may be attributed to the presence of *Ha. leachi*, whose role together with *Ha. paraleachi* as possible vectors of *B. rossi* needs to be investigated. The detection of *T. ovis* in seven dogs is considered unusual.

Of the 80 blood samples from cattle that were tested using the B. bigemina-specific qPCR, 35 (44%) were positive for B. bigemina DNA. The relatively high proportion of B. bigemina infections corresponds with the occurrence of the competent vector, R. decoloratus, the second most frequently collected tick from cattle (90% infestation rate). No B. bovis was detected from cattle using the species-specific qPCR, consistent with the low occurrence of R. microplus. Ehrlichia ruminantium was not detected using RLB, due to its relatively low sensitivity (Byaruhanga et al. 2016). Therefore, qPCR was performed to ascertain infection rates and whether A. pomposum is involved in the transmission. Of the 70 blood samples from cattle, goats, sheep and dogs tested using E. ruminantium-specific pCS20 qPCR, only three tested positive for the pathogen. Of the 100 A. pomposum DNA samples tested, seven were positive to E. ruminantium. Our E. ruminantium detection rate from A. pomposum ticks (7%) is consistent with several other studies in A. hebraeum, A. variegatum and A. lepidum (Muramatsu et al. 2005). This is the first study to determine E. ruminantium infection rates in A. pomposum. Anaplasma platys was detected in cattle (18%) and sheep (6%) and confirmed by 16S rRNA sequence analysis. Of the three near full-length 16S rRNA gene sequences (1249 bp) obtained from three cattle samples, two sequences (MN401149 and MN401150) were 97.8 to 98.0% identical to A. platys sequences from domestic dogs in South Africa (MK814421, query cover 100%) and Cuba (KX792089, query cover 100%) and 98% identical to Anaplasma sp. Omatjenne (U54806). The pathogen has been reported worldwide, mostly from dogs, causing canine cyclic thrombocytopenia. However, variants of A. platys have been implicated as causes of disease in cattle and humans (Sykes and Foley 2014).

The majority of haemoparasites detected in this study are usually associated with the 17 tick species identified, but for some pathogens such as *A. centrale* (usually transmitted by *R. simus*), *E. canis* and *B. vogeli* (usually transmitted by *R. sanguineus*), their usual tick vectors were not found. This points to the possibility that these pathogens may be transmitted by tick vectors other than those which have been previously reported. However, the presence of established biological tick vectors of these pathogens may not be ruled out. A limitation of this study is that ticks were collected from only 20 animals of each host species and only from Tchicala-Tcholoanga Municipality in Huambo Province. Although this may not provide a comprehensive comparison of the spatial and host-specific distribution of tick species in Angola, the findings contribute to the knowledge of the occurrence of ticks and tick-borne pathogen species in the country. The findings provide reference for future research in Angola, involving a larger animal sample and with pathogen DNA sequencing to determine the prevalence, distribution and diversity of the common pathogens.

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Authors' contributions Gourgélia Sili: writing—original draft, investigation, methodology, visualisation, data curation. Charles Byaruhanga: writing—original draft, investigation, visualisation, formal analysis, methodology, data curation. Ivan Horak: resources, writing—review and editing, investigation, validation. Helena Steyn: methodology, investigation. Mamohale Chaisi: methodology, investigation, writing review and editing. Marinda C. Oosthuizen: project administration, conceptualisation, writing—review and editing, Supervision. Luís Neves: project administration, conceptualisation, funding acquisition, writin—review and editing, supervision.

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Data Availability Raw data can be provided on request.

#### **Compliance with ethical standards**

**Conflict of interest** The authors declare that there is no conflict of interest and no competing interests.

**Ethics approval** The research was approved by the Animal Ethics Committee (AEC) of the Faculty of Veterinary Science, University of Pretoria, South Africa (number: V053-16) and followed the AEC guidelines for study design, handling and care of study animals, collection of blood and tick samples and relevant laboratory procedures. Permission to perform the research was granted by the Department of Agriculture, Forestry and Fisheries, Republic of South Africa (Reference number: 12/11/1/1/9).

Consent to participate Not applicable

**Consent for publication** All the authors listed for this publication have agreed on the submission for publication.

Code availability Not applicable

# References

- Byaruhanga C, Collins NE, Knobel D, Chaisi ME, Vorster I, Steyn HC, Oosthuizen MC (2016) Molecular investigation of tick-borne haemoparasite infections among transhumant zebu cattle in Karamoja Region, Uganda. Vet Parasitol Reg Stud Rep 3–4:27–35
- Byaruhanga C, Collins NE, Knobel DL, Khumalo ZTH, Chaisi ME, Oosthuizen MC (2018) Molecular detection and phylogenetic analysis of *Anaplasma marginale* and *Anaplasma centrale* amongst transhumant cattle in north-eastern Uganda. Ticks Tick Borne Dis 9:580–588
- Cardoso L, Oliveira AC, Granada S, Nachum-Biala Y, Gilad M, Lopes AP, Sousa SR, Vilhena H, Baneth G (2016) Molecular investigation of tick-borne pathogens in dogs from Luanda, Angola. Parasit Vectors 9:252
- Dias VS (1948) Subsídios para o estudo dos ixodídeos de Angola, animais domésticos. Anais dos serviços de Veterinária
- Dias JATS (1983) Subsídios para o conhecimento da fauna ixodológica de Angola. Sér Zool 11:57–69
- Durrani AZ, Younus M, Kamal N, Mehmood N, Shakoori AR (2011) Prevalence of ovine *Theileria* species in district Lahore. Pak J Zool 43:57–60
- Gomes AF (1993) The tick vectors of cowdriosis in Angola. Rev élev Méd Vét Pays Trop 46:237–243
- Gomes AF, Neves L (2018) *Rhipicephalus microplus* (Acarina, Ixodidae) in Angola: evidence of its establishment and expansion. Exp Appl Acarol 74:117–122
- Gomes AF, Pombal AM, Venturi L (1994) Observations on cattle ticks in Huíla Province (Angola). Vet Parasitol 51:333–336
- Jongejan F, Uilenberg G (2004) The global importance of ticks. Parasitology 129:S3–S14
- Kim C, Iseki H, Herbas MS, Yokoyama N, Suzuki H, Xuan X, Fujisaki K, Igarashi I (2007) Development of TaqMan-based real-time PCR Assays for diagnostic detection of *Babesia bovis* and *Babesia bigemina*. Am J Trop Med Hyg 77:837–841

- Kubelová M, Mazancova J, Siroký P (2012) *Theileria, Babesia* and *Anaplasma* detected by PCR in Ruminant herds at Bié Province, Angola. Parasite 19:417–422
- Muramatsu Y, Ukegawa S, El Hussein ARM, Rahman MBA, Gabbar KMAA, Chitambo AM, Komiya T, Mwase ET, Morita C, Tamura Y (2005) *Ehrlichia ruminantium*, Sudan. Emerg Infect Dis 11: 1792–1793
- Nijhof AM, Pillay V, Steyl J, Prozesky L, Stoltsz WH, Lawrence JA, Penzhorn BL, Jongejan F (2005) Molecular characterisation of *Theileria* species associated with morality in four species of African antelopes. J Clin Microbiol 43:5907–5911
- Serrano FMH (1963) Considerações sobre morfologia, ecologia e biologia dos ixodideos dos generos *Amblyomma* e d*ermacentor* assinalados em Angola, Revista Ciência Veterinária. Vol. LVIII. N° 386/7
- Sirigireddy KR, Ganta RR (2005) Multiplex detection of *Ehrlichia* and *Anaplasma* species pathogens in peripheral blood by real-time reverse transcriptase-polymerase chain reaction. J Mol Diagn 7:308– 316
- Steyn HC, Pretorius A, McCrindle CME, Steinmann ML, Van Kleef M (2008) A real-time PCR assay for *Ehrlichia ruminantium* using *pCS20*. Vet Microbiol 131:258–265
- Sykes JE, Foley JE (2014) Anaplasmosis, Chapter 29. Canine and Feline Infectious Diseases, pp 290-299
- Thrusfield M (2007) Veterinary epidemiology. Blackwell Science Ltd, Oxford, pp 238–239
- Walker AR, Bouattour A, Camicas J.-L, Estrada-Peña A, Horak IG, Latif AA, Pegram RG, Preston PM (2003) Ticks of domestic animals in Africa: a guide to identification of species. International Consortium on Ticks and Tick Borne Diseases (ICTTD-2), 221 pp

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